Supplementary Materials and Methods.

**Cell culture and transfection**: T cell hybridoma line, DO11.10 and 293T cells were cultured in media containing 50% RPMI and 50% DMEM with 10% fetal bovine serum, 50mM L-glutamine and Gentamycin. 2x10^6 cells of DO11.10 expressing N1^{IC} or empty control were transfected with 4\mu g of NF-\kappaB reporter plasmid and 0.5\mu g of pRL-CMV plasmid as an internal control using the AMAXA nucleofection system according to manufacturer’s protocol for T cells (program S-018). 293T cells were plated at 1x10^6 cells/ml prior to transfection and transfected using Fugene 6 (Roche Co.) following manufacturer’s standard protocols. Cells were processed 48 hours later for luciferase assays, confocal microscopy or western blots.

**Dual-Luciferase assays**: 293T cells were plated on 60mm dishes and transfected with the indicated expression vectors or with empty vector as a control. In the different experiments we used 0.4 \mu g of NF-\kappaBx3 luc as the reporter plasmid and 0.1\mu g pRL-CMV as the internal control. Luciferase assays (Dual-Luciferase Assay System, Promega) were performed 48 hours after transfection, following the manufacturer’s instructions. Luciferase values were normalized against renilla luciferase activity. At least three independent experiments were performed in duplicate.

**Gel shift mobility assays (EMSA)**: Nuclear extracts from splenocytes isolated as described above were subjected to EMSA using the Gel Shift Assay System (Promega, Madison, WI). In some experiments, cells were pretreated with 50\mu M IL-CHO as indicated above. Nuclear extracts were prepared and 5\mu g of extract was incubated with ^32P-labeled NF-\kappaB oligonucleotide (Promega, Madison, WI). For super-shift assays, reactions were incubated for 2 hours in the presence of 2\mu g of indicated antibodies (Santa-Cruz Co.). EMSA and super-shift reactions were resolved by eletrophoresis on a 4% non-denaturing polyacrylamide gel. The gels were dried and subjected to autoradiography.

**Confocal microscopy**: 293T cells were transfected with indicated plasmids and examined 1 day after transfection using laser-scanning confocal microscopy (LSM510, Carl Zeiss) with a 40x phase contrast oil immersion objective (numerical aperture=1.3). Excitation of EGFP was performed using an Argon ion laser at 488 nm. Emitted light was reflected through a 505-530 nm bandpass filter from a 540 nm dichroic mirror.
dsRed fluorescence was excited using a green Helium Neon laser (543 nm) and detected through a 560 nm long-pass filter. Data capture and extraction were performed using manufacturer’s software (Zeiss LSM Image Browser version 3.1).