Subcellular localization of the indicated GFP-tagged proteins associated with 40S pre-ribosomes. For visualization of the nucleolus, GFP-tagged strains (C-terminal, genomic integrations in wild-type strain DS1-2b) were transformed with pRS314-DsRed-NOP1. Cells were also stained with DAPI to visualize DNA. (A) Enp1p, Rrp12p, Dim1p, and Tsr1p have a predominantly nucleolar location, but can also be detected in the cytoplasm. (B) In contrast, Rio2p, Nob1p, and Yor145p localize to the cytoplasm with nuclear exclusion in a representative number of cells.

Fig. SI 1 (Schäfer et al., 2002)
SI 2 A/B (Schäfer et al., 2002)
Fig. SI 2  The *rio2-1* ts mutant encodes a thermolabile *rio2-1p* protein that is dispensable for pre-40S complexes. (A) pRS315-*rio2-1* was C-terminally TAP-tagged and expressed in the *rio2* null strain. Western blot analyses using anti-ProtA antibodies were performed on whole cell lysates derived from cells expressing either Rio2p-TAP or rio2-1p-TAP. Anti-Arc1p antiserum was used to control equal loading. (B) Rio2p-TAP, rio2-1p-TAP, and Tsr1p-TAP were isolated from the indicated yeast strains grown at either 23°C or shifted to 37°C for 4h. Eluates were separated on a 4-12% SDS-polyacrylamide gradient gel and stained with Coomassie. The Rio2p protein is indicated by asterisks. Prominent co-purifying bands are labeled. (C) Subcellular location of Rio2p-GFP and rio2-1p-GFP in wild-type cells (DS1-2b) and the LMB-sensitive *xpo1* -mutant, all grown at the indicated temperatures. (D) Growth properties of strains expressing either Rio2p-GFP or rio2-1p-GFP in *rio2::KAN* as compared to a heterozygous *RIO2/rio2::KAN* strain (wild-type). Serial dilutions were spotted on YPD plates and incubated for 3 days at 23°C, 30°C, and 37°C.
Fig. SI 3 Sedimentation behavior of the Rps2p-eGFP reporter construct expressed in *enp1-1* and *rio2-1* ts mutants. Both strains were transformed with pRPS2-eGFP, propagated at either permissive (23°C) or restrictive (37°C) temperatures and subjected to sucrose gradient centrifugation. Rsp2p-GFP was visualized by Western blot analysis of the collected gradient fractions using an anti-GFP antibody.